ORIGINAL ARTICLES

In vivo the rapeutic effects of albendazole in combination with IFN- α on cystic echinococcos in mice

Zhelin Yun*1, Yongsheng Zhou1, Yan Zhang2, Bing Li1

¹Department of General Surgery, Baogang Hospital, Baotou, Inner Mongolia, China ²The Sci-technology Center, Ningxia Medical University, Yinchuan, Ningxia, China

Received: August 30, 2018	Accepted: October 18, 2018	Online Published: December 10, 2018
DOI: 10.14725/dcc.v5n4p9	URL: http://dx.doi.org/10.14725/	dcc.v5n4p9

Abstract

Objective: This study was aimed to evaluate the in vivo therapeutic effects of the combination of ABZ with interferon (IFN)- α on cystic echinococcosis (CE) in mice.

Methods: After 5-month secondary infection with protoscolices, Balb/c mice were randomly divided into four groups: ABZ-treated group, IFN- α -treated group, ABZ+IFN- α group and untreated control group. Drugs in different treated groups were respectively administered for 2 months. Blood was respectively taken from caudal veins in mice at treatment time points of 0 d, 7 d, 14 d, 28 d, 36 d, 48 d and 60 d, in order to detect the changes in the level of antibodies in the serum. Mice were then sacrificed after the end of treatment, and related indicators were detected to evaluate the therapeutic effects.

Results: In comparison with untreated control group (p < .01) and ABZ-treated group (p < .05), ABZ+IFN- α group showed a more significant decrease in the number of cysts, cyst size and cyst weight. After transmission electron microscopy (TEM) of the cysts in different treated groups, it was found that, there was an obvious change in cyst ultrastructure in ABZ+IFN- α group. ELISA experimental results showed that, in ABZ+IFN- α group, there was a significant decrease in IL-10 in serums and splenocytes (p < .01); In comparison with untreated control group, ABZ+IFN- α group showed a more significant decrease in the levels of IgE, IgG and its subtypes (p < .01).

Conclusions: In this study, the combination of ABZ with IFN- α was proved to be an effective CE treatment option.

Key Words: Cystic echinococcosis, Albendazole, IFN- α , IL-10, Ultrastructural modification

Cystic echinococcosis (CE) is a worm-like zoonosis that is widely spread throughout the world and even highly prevalent in some regions.^[1] China is one of the countries with the highest incidence of echinococcosis.^[2] The treatment of CE usually depends on surgery and/or chemotherapy, and the option hinges on a variety of factors such as the size and location of the cyst, accompanying microbial infections, potentially dangerous complications (e.g. cyst rupture, etc.)^[3–6] As a supplement to surgical treatment, the only available drug for CE is benzimidazole carbamate derivatives, namely mebendazole (MBZ) and albendazole (ABZ).^[7] However, more than 20% of CE cases responded poorly to the treatment.^[8] Therefore, there is an urgent need for some novel and more effective treatment options.

^{*}Correspondence: Zhelin Yun; E-mail: Yunzhelin727@126.com; Address: Department of General Surgery, Baogang Hospital, Baotou, Inner Mongolia, China.

Interferon (IFN)- α , a type of Grade 1 interferons, was originally used mainly for antiviral treatment of various viral diseases.^[9] Subsequently, IFN- α was confirmed to have a variety of biological functions, including: antiviral function, anti-tumor function and immune regulation etc.^[9,10] For another type of echinococcosis, i.e., alveolar echinococcosis (AE), some studies have shown that IFN- α has positive protective effects. Godot V et al. have confirmed that IFN- α can significantly reduce the cysts in patients with AE.^[11] It has been further found in the studies that IFN- α can reverse Th2-type responses to Th1-type responses during the treatment of AE, and it is beneficial for body immunity to provoke an effective protective immune response to the cysts. In the study concerning AE, it was also found that IFN- α can relieve the degree of liver fibrosis and injury. However, currently, there is a lack of researches about the therapeutic effect of IFN- α on CE. This study was designed to evaluate the in-vivo the rapeutic effect of IFN- α and ABZ-IFN- α on CE in mice.

1 Materials and methods

1.1 Materials

1.1.1 Experimental animals

All animal experiments were approved by the Ethics Committee in Baogang Hospital in Inner Mongolia. Forty Balb/c female mice (8 weeks old, 18-20 g) were purchased from Beijing Vital River Laboratory Animal Technology Co., Ltd. The experimental animals were raised in a controlled environment at room temperature $(22^{\circ}C \pm 1^{\circ}C)$ under a certain lighting condition (12 h light/dark cycle).

1.1.2 Drugs and reagents

ABZ was purchased from GSK, and Pegasys® interferon alfa-2a was purchased from Roche. Penicillin, streptomycin, RPMI 1640 cell culture medium and fetal bovine serum (FBS) were all purchased from Invitrogen. Con-A was purchased from Sigma-Aldrich. ELISA assay kits for mouse IL-4, IL-10 and IFN- γ cytokines were purchased from e-Bioscience. Goat Anti-Mouse IgE, IgG antibodies and its subtypes of the second antibody were purchased from Novagen. Other reagents were of domestic analytical purity.

1.1.3 Protoscolices

After the cysts were isolated with the help of clinical surgery, they were quickly and aseptically transferred to the laboratory. The cystic fluid was extracted from the cysts by use of a syringe, with the weight of $9,000 \times g$, and centrifuged at 4°C for 20 min, with the supernatant discarded.

The settled protoscolices were washed twice by use of PBS containing double antibody (pH 7.2, containing 1,000 μ g/ml penicillin and 1,000 U/ml streptomyces). The activity of protoscolices was identified with the help of Meilan exclusion experiment.^[12] Only when the activity of protoscolices reaches 90% or more will it be used for subsequent secondary infection in mice. The final concentration of protoscolices used for secondary infection was 1,500/200 μ l PBS.

1.2 Experimental methods

1.2.1 Experimental design

40 Balb/c female mice got secondary infection by means of intraperitoneal injection, each mouse was infected with 1,500 fresh protoscolices resuspended in 200 μ l of PBS. In 5 months after infection, these mice were randomly divided into four groups (10 mice/group): (1) ABZ-treated group, in which mice were given ABZ (5 mg/kg) by gavage; (2) IFN- α -treated group, in which mice were given intramuscular injection of IFN- α -2a (10,000 IU/50 μ l) every 3 d; (3) ABZ+IFN- α -treated group, in which mice were given ABZ and IFN- α , with the same dosage as in (1) and (2); (4) Untreated control group. The drug administration in each group was maintained for 2 months. Subsequently, the mice were sacrificed, and related indicators were detected to evaluate the therapeutic effect.

1.2.2 Detection of parasitic infection and analysis of effective rate of treatment

The abdominal cavity of each mouse was dissected to measure the number of cysts, cyst size and cyst weight. Effective rate of treatment (based on cyst weight of the infected mouse) was calculated by the following formula:^[13]

Effective rate = $(Xc - Xt)/Xc \times 100\%$

Whereas, Xc is the average cyst weight of untreated control group, and Xt is the average cyst weight of each treated group.

1.2.3 Transmission electron microscopy (TEM)

After each group of cyst samples were processed, they were detected by use of TEM.^[14] The cysts were cut into blocks the size of 1 mm³, and fixed with the pre-cooling 0.1 M PBS (containing 2.5% glutaric dialdehyde) for 5 h. These samples were washed twice with 0.1 M PBS, and fixed with 1% osmium (VIII) oxide. The samples were then dehydrated in an ascending series of ethanol concentrations and embedded overnight in an equal volume of propylene oxide/epoxy resin. Subsequently, it was again embedded in 100% fresh epoxy resin for 24 h and at 65°C for 18 h. Ultramicrotome

(Leica EM FC7) was used to prepare sections with the thickness of 50-70 nm. Afterwards, these sections were placed on 200-mesh copper sieve, and got immersed in saturated uranium acetate and lead citrate staining solutions for 10 min and 5 min respectively. The sections were washed with distilled water and dried naturally. TEM (H7650, Hitachi) was applied to the measurement.

1.2.4 Enzyme linked immunosorbent assay (ELISA)

First of all, splenocytes isolated from each mouse were resuspended by use of RPMI 1640 medium (containing 10% heat-inactived FBS), with the final concentration adjusted to 5×10^6 splenocytes/ml, added by 2.5 µg/ml Con-A (Sigma) stimulants. Under the condition of 37°C and 5% CO₂, these cells were incubated for 72 h, followed by the collection of culture supernatant to detect the concentrations of IL-4, IL-10 and IFN- γ . Specific procedures for the detection of cytokines in serum and cell culture supernatant are described in the instructions of EILSA Cytokine Assay Kit.

ELISA was used to detect the level of antibodies in the serum: 96-well microplate was used in this experiment, 100 μ l of 0.1 M phosphate buffer (pH 9.6, containing 0.1 μ g/ μ l cyst crude antigen) was added into each well to be incubated overnight at 4°C. The serums were diluted into PBST buffer (pH 7.2, with PBS containing 0.05% tween-20) under the ratio of 1:100, and incubated at 37°C for 1 h. Subsequently, the second antibody, i.e., HRP-marked Goat Anti-Mouse IgG, IgG subtype and IgE under the ratio of 1:1,000, was added. OD value of A490 nm was measured by the enzyme-labeled instrument (Bio-Rad).

1.3 Statistical analysis

All data were processed by Prism 5.0 (GraphPad Software) for analysis. The results were represented by mean \pm standard deviation. Cyst weight, size, the concentration of cytokines and the level of antibodies were analyzed by one-way ANOVA respectively. The difference p < .05 was considered to be statistically significant.

2 Results

2.1 ABZ+IFN- α can significantly suppress the cysts

All the mice were not dead during the course of the experiment, indicating that there were no serious side effects of drugs. When these mice were sacrificed, the difference in the average weight among 4 groups of mice was of no statistical significance (p > .05). The number of the cysts was obviously lower in ABZ+IFN- α -treated group than untreated control group (p < .01) or ABZ-treated group (p < .05); in comparison with untreated control group, the number of the cysts was obviously decreased in ABZ-treated group (p < .01). However, there was no significant difference in the number of the cysts between IFN- α -treated group and untreated control group (p > .05, see Table 1).

The results showed that, not only cyst weight but also cyst size was obviously decreased in ABZ+IFN- α -treated group in comparison with untreated control group (p < .01) and ABZ-treated group (p < .05); in ABZ+IFN- α -treated group, the inhibition rate of the cysts reached 89.7%. It was indicated that the combination of ABZ and IFN- α had a more significant therapeutic effect (see Table 2).

 Table 1: The number of the cysts in untreated control group and different treated groups

Group	The number of the cysts in each mouse Mean ±SEM					Mean ±SEM					
ABZ group	3	2	2	1	4	1	2	1	3	2	2.100 ± 0.314
IFN-α group	4	7	3	4	5	1	6	2	4	2	3.800 ± 0.592
ABZ+IFN-α group	1	1	1	0	2	1	1	2	0	0	$0.900\ \pm 0.233^{*\#}$
Untreated control group	5	4	0	6	2	3	3	5	7	4	3.900 ± 0.640

Note. The number of the cysts was more significantly decreased in ABZ+IFN- α -treated group than untreated control group (*p < .01) and ABZ-treated group (*p < .05)

Table 2:	Cyst weight	and size in	different groups

Tuble = Cyst weight and size in anterent groups						
Group	Cyst weight (g)	Cyst size (mm)	Inhibition rate of the cysts (%)			
ABZ group	$0.21 \pm 0.17^{*}$	$2.1 \pm 0.9^{*}$	78.4			
IFN-α group	0.89 ± 0.28	6.7 ± 1.5	0.08			
ABZ+IFN-α group	$0.10\ {\pm}0.05^{*\#}$	$1.2 \pm 0.4^{*\#}$	89.7			
Untreated control group	0.97 ± 0.45	7.3 ± 3.2	-			

Note. Cyst weight and cyst size was more significantly decreased in ABZ+IFN- α -treated group than untreated control group (*p < .01) and ABZ-treated group (*p < .05)

2.2 Ultrastructural modification and damages of the cysts in ABZ+IFN-α-treated group

TEM was used to observe ultrastructural modification of the cysts in different groups. The results indicated that the cysts in untreated control group showed a typical cystic structure of echinococcus granulosus, while ABZ+IFN- α -treated group showed ultrastructural modification and damages (see Figure 1A and 1B). In the germinal layer, normal undifferentiated cells, fibrous tissues, typical nuclei and nucleolus structures in the cysts of mice in ABZ+IFN- α -treated group

disappeared, and karyorrhexis and a large number of lipid droplets appeared instead. As was shown in Figure 1C and 1D, it indicated that the ability of the germinal layer to produce daughter cysts had been damaged. In comparison with untreated control group, the typical cortex and microtrichia microtubule structures of the mouse cysts disappeared in ABZ+IFN- α -treated group (see Figure 1E and 1F). The non-cellular, sugar-rich corneum of the normal outer layer was replaced by an irregular and vacuole-rich structure (see Figure 1G and 1H).



Figure 1: Ultrastructural modification of the cysts in ABZ+IFN- α -treated group ABZ+IFN- α treated group (A, C, E and G) and untreated control group (B, D, F and H). A and B showed the general ultrastructure of the cysts; C and D showed the germinal layer of cysts; E and F emphasized on the changes of teguments and microtriches; F was derived from red rectangle area of D; G and H represented the laminated layer of cysts. In untreated control group, the cyst has normal laminated layer (LL), germinal layer (GL), tegument (Te), microtriches (Mr), undifferentiated cells (UC), fibrous tissue (F), cell nucleus (Nu) and nucleolus (Nl). In ABZ+IFN- α treated group, ultrastructural modification shows as lipid droplets (Ld) and vaeuloles (V)

2.3 ABZ+IFN-α treatment can significantly reduce the level of IL-10 in the serum

To detect the formation of cytokines after treatment, it was found through the detection that the level of IL-10 in the serum was obviously decreased in ABZ+IFN- α -treated group than untreated control group (2.29 ± 0.25 ng/ml vs. 3.44 ± 0.14 ng/ml, p < .01) (see Figure 2A). There was no significant difference in the levels of IFN- γ (see Figure 2B, p = .08) and IL-4 (see Figure 2C, p = .11) among different groups.

2.4 ABZ+IFN-α treatment can significantly reduce the level of IL-10 secreted by splenocytes

After detection of the ability of splenocytes to secrete cytokines, it was found that the level of IL-10 was obviously decreased in ABZ+IFN- α -treated group than untreated control group (0.91 ± 0.10 ng/ml vs. 1.99 ± 0.17 ng/ml, p < .01) (see Figure 3A). However, there was no significant difference in the levels of IFN- γ (see Figure 3B, p = .31) and IL-4 (see Figure 3C, p = .18).

2.5 The changes of specific antibodies in the serum in ABZ+IFN-α-treated group

In order to better understand the effect of this treatment option on the level of specific antibodies, ELISA was performed to detect the levels of serum antibodies in different groups. The results showed that, in comparison with untreated control group, the level of IgG antibody was being decreased in ABZ+ IFN- α -treated group, and obviously decreased at the end of the treatment (on 60th d) (p < .0001, see Figure 4A). In addition, at the end of the treatment, the levels of the subtypes of IgG antibody, including IgG1 (p < .0001, see Figure 4B), IgG2 (p = .0299, see Figure 4C) and

IgG4 (p < .0001, see Figure 4D) had been significantly decreased as the treatment progressed. Meanwhile, the level of

IgE was also significantly decreased in ABZ+IFN- α -treated group (p < .0001, see Figure 4E).



Figure 2: Significant decrease of IL-10 in the serum in ABZ+IFN- α -treated group ELISA assay kit was used to detect the concentrations of IL-10 (A), IFN- γ (B) and IL-4 (C) in the mouse serums in different groups; **p < .01



Figure 3: Significant decrease of IL-10 secreted by splenocytes in ABZ+IFN- α -treated group ELISA kit was used to detect the concentrations of IL-10 (A), IFN- γ (B) and IL-4 (C) in the supernatant of mouse splenocytes in different groups; **p < .01



Figure 4: The detection of IgG, IgG1, IgG2, IgG4 and IgE

3 Discussion

The therapeutic effect of ABZ in combination with IFN- α on CE was evaluated in this study. The results showed that ABZ combined with IFN- α had a better therapeutic effect on mice with secondary infection of CE. It was further confirmed that the therapeutic effect may be related to ultrastructural modification of the cysts, decreased IL-10 level and lowered humoral immunity.

IFN- α has a variety of biological functions including antiviral infection, anti-tumor and immune regulation, but the research on the role in helminthic infection is very limited in its amount. However, it is worth mentioning that in the study of AE, IFN- α can relieve cystic lesions in humans and mice.^[11,15] The researchers further stated that this type of effect could be related to the reverse of Th2-type cytokines to Th1-type cytokines, including the down-regulation of IL-10, IL-6 and IL-13 and the up-regulation of IFN- γ .^[11] However, this study showed that only IFN- α had no effect on alleviating CE or reversing the type of Th cytokines, which indicated that only IFN- α could not break the Th2-type immune response of chronic CE. This result was different from previous reports on AE treatment. However, IFN- α combined with ABZ can significantly relieve CE to reduce the burden of the cysts. Compared with ABZ-treated group or untreated control group, ABZ+IFN- α -treated group has a better therapeutic effect (see Tables 1 and 2). Further studies have shown that this combination therapy can significantly down-regulate IL-10. IL-10 is a typical Th2-type cytokine that inhibits the Th1 protective response in CE and helps parasites evade the host immune response.^[16] Therefore, the better therapeutic effect of the combination therapy may be related to the inhibition of IL-10 and the reduction of Th2-type immune response.

The humoral immune response plays an important role in the occurrence and development of CE. In the early stage of infection, it develops an important protective immune response through complement activation, opsonization and ADCC. Among the antibodies in the chronic phase of infection, especially the IgG4 type, has an effect in facilitating the escape of parasites.^[17,18] IgG, IgG1, IgG2, IgG4 and IgE were all significantly decreased in the combination therapy of this study, indicating that the abnormal rise of humoral immune response in the chronic phase of infection was significantly down-regulated during the process of the

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treatment, with IgG4 also significantly reduced. However, although the level of IgG was significantly reduced after the combination therapy, it was found that the level was still higher than the negative level. This may be related to the immune memory of the infection, but it remains to be further studied. In conclusion, the therapeutic effect of the combination therapy may also be related to the down-regulation of the humoral immunity beneficial to the parasites.

In this study, ABZ+IFN- α or IFN- α alone did not significantly increase the level of IFN- γ in the chronic phase of infection. This may be related to the fact that IFN- γ often plays an important protective role in the early stage of infection (7-8 d) and then decreases in the chronic phase of infection.^[19] The effect of the early treatment of infection on IFN- γ will be further investigated. In addition, some studies have reported that IFN- α has an anti-tumor effect by enhancing the antigen presenting function of dendritic cells (DCs),^[20,21] but it remains to be further explored that whether DCs play a role in this combination therapy.

In conclusion, this study confirmed that ABZ combined with IFN- α has a better therapeutic effect on CE and provides an experimental basis for CE treatment.

Conflicts of Interest Disclosure

The authors have no conflicts of interest related to this article.

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