Intrathoracic Nocardia-abscess as a cause of haemodynamic instability

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Abstract
A 64-year old patient was admitted to our hospital one year after renal transplantation. He complained about headache, cough and fever. Blood cultures revealed Nocardia nova. Because of disseminated disease, he was treated with both co-trimoxazol and meropenem. Eight days later, his condition worsened. A CT-thorax was performed which showed large mediastinal masses and compression of vena cava superior and right atrium. The same evening, he underwent a thoracotomy because of haemodynamic instability. The aspirated pus again showed Nocardia nova. He recovered but developed pleural effusion and again this was positive for Nocardia nova. Both meronem and co-trimoxazol were continued. Six months after diagnosis, he was admitted because of gastro-intestinal bleeding and a mycobacterium species was found in gastric and duodenal biopsies. He was started on claritromycine and myambutol, but developed renal failure. Ten months after diagnosis of disseminated Nocardiosis, he died because of worsening condition and renal failure. No post-mortem examination was performed.

Revision of literature showed no other reports of hemodynamic instability by Nocardia-abscesses or compression on vena cava or right atrium.

Keywords
Nocardiosis, Renal transplantation, Nocardia abscess, Mediastinal masses

1 Introduction
Nocardia spp is a ubiquitous gram positive bacteria, which causes localised or disseminated diseases, especially in the immunocompromised. It is an infrequent infection, but incidence is rising, partly due to an increase in immunosuppressed patients for example after transplantation. Nocardiosis occurs worldwide: the incidence might be underestimated because of nonspecific complaints and delay in diagnosis can easily occur. This is worsening the prognosis for an illness that already has a high mortality rate.

2 Case report
A 64-year-old patient was admitted one year after non-heart-beating kidney transplantation because of worsening kidney function without evident cause. His medical history revealed haemodialysis and PCI after myocardial posterior infarction. The latest echocardiogram showed an aortic valve stenosis, gradient 36mmHg.
At the time of admission, he still used a triple immunosuppressive regimen -prednisone 10 mg a day, tacrolimus (3-6 u/l) and mycofenolate mofetil--; he also used metoprolol, diuretics, monocedocard, ascal and a statine. Signs of rejection had not occurred and he had been using co-trimoxazol prophylaxis for six months after transplantation.

At admission, he complained about a severe right-sided headache, fever, shortness of breath, tiredness and night sweats, which existed for at least five weeks. He did not cough but shortness of breath at exertion existed; he only produced a small amount of uncoloured sputum. He did not have noticed differences in urine of micturation.

On physical examination he appeared moderately ill, temperature 38.3°C, blood pressure 120/70 mmHg, pulse rate 108/min. A loud, already known systolic murmur was heard, grade III-IV/VI and also diffuse rhonchi at the right hemithorax. No further abnormalities were found, especially no stigmata of infective endocarditis.

Laboratory results showed leucocytosis (18.1/nl, normal value 4-10/nl) and an elevated C-reactive protein (199 mg/L, normal value < 5 mg/L). Kidney function had mildly deteriorated, serum creatinin 165 umol/l, liver function was without abnormalities. An ECG showed a known right bundle branch block; chest-X-ray showed no signs of pneumonia and sinus radiography was without abnormalities (see Figure 1). Viral swabs remained negative, including on H1N1. Also mastoiditis was ruled out. Although he did not did not reveal signs of sepsis of urinary origin, he was started on ciprofloxacin and cefuroxim.

Within 36 hours, both blood- and urine cultures were found positive with Enterobacter species. Because of persisting periods of fever, more blood cultures were drawn and incubated in a BacT alert system (Biomerieux, France). After 4 days of incubation the blood cultures became positive with branched Gram-positive rods. Because of the Gram stain and the aspect on the blood agar, this micro-organism was suspected to be a Nocardia species. For identification at the genus species level a PCR of the HSP 65 was used [1]. The gen product was thereafter sequenced as described by Rodrigues-Nava [2]. The isolate appeared to be a Nocardia nova. Susceptibility testing was performed by E-test (The E-test, AB Biodisk, Solna, Sweden). The micro-organism was susceptible for co-trimoxazole (MIC 0.075 mg/L), meropenem (MIC 0.0032 mg/L), clarithromycin (MIC 0.032 mg/L), ceftriaxone (MIC 0.5 mg/L), linezolid (MIC 0.075 mg/L), and amikacin (MIC 0.125 mg/L) and resistant to moxifloxacin(MIC 2 mg/L), amoxicillin/ clavulanic acid (MIC > 256 mg/L), and minocycline (MIC 4 mg/L).

**Figure 1.** Thoracic X-ray: enlargement mediastinum.
Immunosuppression was reduced, mycofenolate mofetil was discontinued and antibiotic therapy was switched to meropenem 500 mg 3 times daily. Lumbar puncture showed no signs compatible with meningitis (white blood count 1*10^6/L, protein 0.26 g/L, glucose 4.4 mmol/L). CT cerebrum and culture of cerebral spinal fluid were negative. His headache, of which he had been complaining for 4 weeks, disappeared with the antimicrobial regimen.

One week after change in antibiotic regime, the patient suddenly became hypotensive with persisting tachycardia, complaining about dyspnoea and chest pain. ECG showed no signs of myocardial ischemia or infarction. A transoesophageal echocardiogram was performed, showing no signs of endocarditis, nor valve-destruction or pericardial fluid. CT-thorax was performed mainly to rule out massive pulmonary embolism. This revealed a large intrathoracic mass, with compression of vena cava superior and right atrium, suspicious for malignancy (see Figure 2). However, in view of the positive nocardia cultures, pulmonary abscesses were suspected and a thoracotomy was performed. A large amount of pus was evacuated in which also Nocardia nova was detected. Also a culture of sputum revealed Nocardia. Intravenous meropenem was continued and combined with orally administrated co-trimoxazol. After an initial short period of improvement, his condition again worsened, mainly because of progressive shortness of breath. A second CT-thorax showed a large amount of pleural fluid at the right hemithorax. Diagnostic pleural puncture exhibited a highly viscous exsudative fluid. Because of this viscous nature, a re-thoracotomy had to be performed. Nocardia did not grow in fluid-culture. Following this procedure, the patient recovered and he was planned for percutaneous aorta valve-replacement as soon as he would recover the nocardial infection.

Six months after recovery, the patient was admitted again for repeatedly gastro-intestinal bleeding. Neither gastroduodenoscopy nor coloscopy revealed a bleeding location, but a rare aspect of duodenal mucosa was found. Biopsies were performed, which became positive for a Mycobacterium species, specified as a Mycobacterium genavense (PCR and culture were performed at the Regional Public Health laboratory).

The patient was started on claritromycine and myambutol. Along with diagnosis of this Mycobacterium and after starting this treatment, unfortunately his kidney function detoriated and he developed transplant failure. Because of worsening condition, he was not started on haemodialysis. Two weeks after start treatment for Mycobacterium and almost ten months after diagnosis of disseminated Nocardiosis, he died; unfortunately, no postmortal examination was performed.

3 Discussion
This case report describes clinical deterioration and hemodynamic instability because of compression of vena cava superior and right atrium by Nocardia nova abscesses. Review of the literature showed only four other cases of vena cava
superior syndrome caused by different species of Nocardia \cite{3-6}. Like in our case, in all four patients malignancy was also considered, sometimes delaying diagnosis and treatment. In two cases, drainage was needed to evacuate the effusion, even though the patients were already treated with intravenous antibiotics. One patient, who was allergic to sulphamamide and therefore was treated with streptomycin and erythromycin, died because of ongoing infection and intraspinal dissemination combined with renal failure. This latest was caused by the septic condition, underlying prior condition (systemic lupus) and nephrotoxic medication, one of which being the necessary antibiotics \cite{3}.

Our patient died of a co-infection with M. genavense, a rare nontuberculous mycobacterium, which is mostly found in patients with inherent or acquired immunodeficiency. Recent reports state that these mycobacteria are increasingly being recognized in solid organ transplant recipients, but because of their diversity, therapy is complex and requires a combination of active antimicrobial agents \cite{7,8}. In our case, the actual determination and antimicrobial susceptibility guided our treatment, but soon after start of the combined antimicrobial agents, the patient developed transplant failure and died shortly after.

Nocardiosis is a localized or disseminated infection caused by aerobic actinomycete-like bacteria particularly in immunocompromised patients. Its first descriptions in 1888 were made by Edmund Nocard who believed it to be a bovine pathogen \cite{9,10}. In 1893 Eppinger described the first human case as a pseudotuberculous syndrome in pulmonary disease combined with brain abscesses \cite{11}. Initially this species was thought to be rare, but nowadays, it’s obvious these bacteria are responsible for a wide spectrum of diseases, especially in patients with abnormal immune systems \cite{9,12}. Of the nearly 70 species of the genus Nocardia, over 50 have been reported as causing human infection \cite{13,14}.

Human-to-human transmission has not been documented \cite{10,15}. In most cases, infection is acquired by inhalation of fragmented bacterial mycelia, but transcutaneous inoculation occurs as well as inoculation of the eye \cite{9,10,13}. Virulence of Nocardia spp depends upon this route of administration and inoculums, but also of growth rate and strain \cite{17}. Natural resistance to infection seems to be high, since frequent contact with the organism in the environment probably occurs \cite{17}.

In the US, 1000 cases of Nocardiosis annually are reported, 85\% of which is pulmonary or systemic \cite{9,18}. It is estimated that at least 50\% of patients with Nocardia species infections have underlying immune compromise \cite{9,10,18,19} and only less than 10\% of patients has no definable underlying predisposing factor \cite{17}. Incidence rates after solid organ transplantation differs between 0.7\%-4.0\% and almost primarily described after renal or heart transplantation \cite{16,20,21}.

The time after transplantation to onset of Nocardiosis ranges from 1 week tot 338 weeks, with an average interval of 30 weeks \cite{11}. Thus, the first year after transplantation carries the greatest risk and is mostly associated with episodes of allograft rejection (treated with high-dose steroids) and chronic viral infections, especially the presence of immunomodulating viruses like cytomegalovirus. The timeframe after a period of acute rejection to Nocardia infection averages sixteen weeks \cite{12,16,17}.

In over 60\% of cases, the lungs are the primary site of Nocardia infection. Clinical symptoms include cough, night sweats and fever appearing usually with a subacute pneumonia \cite{9,10,15}. Chest X-ray may be normal, but can also include bilateral disease and cavitations in which necrosis leads to the formation of abscesses \cite{12,14,16}. As a result, multiple radiographic findings have been demonstrated in nocardiosis, including single or multiple nodules, lung masses [with or without cavitations], reticulonodular infiltrates, interstitial infiltrates, lobar consolidation, subpleural plaques, and pleural effusions. Therefore, nocardiosis has frequently been misdiagnosed initially as tuberculosis [since upper lobe involvement is also common and Nocardia spp are weakly acid-fast], invasive fungal disease, and malignancy \cite{20}.

Concomitant pleural effusion occurs in 10\%-30\% of cases and bronchopleural fistulae may develop spontaneously \cite{9}. Initial pulmonary lesions may lead to haematogenous dissemination \cite{12,18}. Fever, anorexia, weight loss and malaise are common and remission as well as exacerbations are frequent over periods of several weeks \cite{9,10}. In case of cerebral involvement, headache, lethargy or altered state of consciousness has been reported. Seizures or focal neurological
abnormalities may indicate focal brain infection [20]. Occasional patients present with subacute or chronic meningitis and
during further evaluation brain abscesses are usually found. The combination of headache and unexplained fever in
immunosuppressed patients should therefore raises suspicion and necessitates a complete and urgent neurological
evaluation by CT of the head and lumbar puncture [20].

We could not diagnose Nocardia in cerebral spinal fluid nor find abnormalities at CT cerebrum in our patient. However, it
is known from literature that CNS involvement may occur with gradual or sudden onset of headache and because the
headache disappeared as soon as we started the treatment for Nocardia, we could not rule out brain involvement.

Although dissemination is presumed to be via haematogenous spread, capture of Nocardia in blood cultures is
unusual [9, 16]; routine cultures usually remain negative. Co-infection with more resilient micro-organisms [which
frequently exists] or intermittent and infrequent bacteraemia could possibly explain this phenomenon [20]. Furthermore,
Nocardia spp is a relatively slow-growing organism which prefers enriched media [16]. Even on this specific media, culture
usually may require 4-21 days to develop characteristic appearance [9, 11]. Thus, to optimize recognition, the microbiology
laboratory should be notified when Nocardia is suspected [9, 10, 15].

Nocardia spp is best examined in cultures of sputum, bronchoalveolar lavage fluid or pus samples from abscess or wound
drainages [9, 10, 16, 18]. Especially in refractory cases, susceptibility testing should be considered, because Nocardia species
can vary in their antimicrobial susceptibility patterns [21].

The standard therapy of Nocardiosis consists of prolonged administrations of trimethoprim - sulfamethoxazole
(co-trimoxazole). Co-trimoxazole is available in intravenous and in oral form and has excellent pharmacokinetic
properties including crossing the blood - barrier barrier [9, 10, 15]. The in vitro susceptibility testing of Nocardia spp has a
good correlation with the reducing of colony counts of experimental models [22]. Despite the use of in vitro sensitivity data
to guide therapy, failures occur, often secondary to poor immune status of the host or resistance to several antimicrobial
agents [9].

Alternatives to the standard regimen includes carbapenems, ampicilline, minocycline [9, 10, 12]. For some Nocardia species,
third generation cephalosporin may be used, but N. farcinica one the most frequent isolated species is relatively resistance
to these antibiomicrobial agents. Linezolid is promising in the treatment of Nocardia infections, but because of toxic side
effects this drug is not recommended for long-term therapy.

According to the majority of reports, also in case of disseminated infections a combination of two different drugs are
advocated. In an immunocompromised mice-model, imipenem and amikacine appeared to be the most bactericidal
regime [22], but due to its poor CNS penetration, this combination does not provide optimal therapy. Another carbapenem,
i.e. meropenem, is often used instead of imipenem because of the minor chance of convulsions. As in our patient,
abscesses or purulent effusions should be treated with aspiration and/ or drainage if possible, especially if these are large
and does not respond sufficiently to antibiotics [9, 16, 19].

Nocardia-infections tend to relapse so long courses of antimicrobial therapy are often needed [20]. Due to the lack of
comparative clinical studies, the optimal therapy for Nocardiosis is not known especially since time to relapse ranges from
one month to several years after previous infection [10, 15, 19]. Treatment duration of at least six months is recommended
for immunocompromised patients and therapy in CNS nocardiosis or extensive infection should be at least continue for 12
months [9, 12, 15].

Worldwide, co-trimoxazol prophylaxis for pneumocystis jirovecii is recommended in solid organ transplantations at least
for six months after transplantation [17]. This prophylactic regime also gives protection against other opportunistic
infections such as toxoplasmosis and listeriosis. However, this low dose prophylaxis is not a guarantee for absolute
protection against Nocardia infection despite favourable susceptibility results [9].
Due to the long lasting course of infection and the risk of dissemination or formation of abscesses, mortality of pulmonary or systemic nocardiosis is estimated at 15%-40%, but involvement of CNS increases the mortality [10]. Overall mortality rate is 44%-85%, depending on the underlying immune status of the host [9, 14, 19]. It is estimated that 10% of the infectious death after organ transplant is due to Nocardiosis [9].

4 Conclusion
Nocardia spp is an opportunistic pathogen, causing a spectrum of diseases ranging from non-specific subacute pneumonia to formation of pulmonary abscesses. As the presenting symptoms are non-specific, there is usually a delay in diagnosis. It is important not to misinterpret any pulmonary lesion and to suspect opportunistic diseases like Nocardiosis, especially in immunosuppressed patients who develop pulmonary symptoms [14].

Even for seemingly minor findings CT or MRI of lungs, abdomen or brain should be considered, especially when disseminated disease is suspected [9, 12, 18].

Therapy of choice is co-trimoxazol, although animal studies have shown that imipenem combined with amikacyn may be the best opportunity. Duration of therapy should be long, especially in disseminated disease and exacerbations. Complications such as empyema, mediastinitis, pericarditis, and superior vena cava syndrome can occur following contiguous spread of nocardial infection from a lung, pleural, or cutaneous focus [3, 4, 23].

Mortality rates are high, despite long courses of treatment. Our clinical case describes a patient who became haemodynamic unstable because of compression of vena cava superior and right atrium and needed an urgent operation to be stabilized. By our knowledge, there are only a few cases described in literature with an infection to this extent.

References


